

## **Cloning and Expression Vectors**

| CATALO                   | G  |  | CATALOG<br>NUMBER  |               |
|--------------------------|--|--|--|---------------|
| 101084<br>0°C            | RIBONUCLEASE B<br>From Beef Pancreas<br>Activity: 50 Kunitz units/mg.<br>Salt and protease free<br>Prep'd from cryst. RNase  | 50 mg<br>100 mg<br>500 mg<br>1 g   | <ul> <li>T7 RNA POLYMERASE<br/>Isolated from <i>E. coli</i></li> <li>The enzyme is very specific for T7 promoters. These<br/>promoter sequences, when inserted into cloning vectors,<br/>can be used to synthesize RNA transcripts of the cloned<br/>DNA sequences. These transcripts are useful as<br/>hybridization probes and substrates for RNA systems.</li> <li>Unit Definition: One unit catalyzes the incorporation of 1<br/>nmol of labeled ribonucleotide into acid precipitable<br/>material in one hour at 37°C.</li> <li>Shipping &amp; Storage: Solution in 20 mM potassium<br/>phosphate, pH 7.5, 0.1 mM EDTA, 0.1 mM DTT and 50%<br/>glycerol.</li> <li>Shipped on dry ice.</li> </ul> | 5 KU          |
| 104907<br>0°C            | RIBONUCLEASE B<br>Lyophilized; Phosphate Free<br>Essentially protease-free<br>Activity: Approx. 100 Kunitz units/mg  | 10 mg<br>100 mg<br>500 mg  |  |               |
|                          |  |  | Expedite Your T7/SP6 Transcription With ICN.   |               |
| 152025<br>0°C            | <b>RIBONUCLEASE H</b><br>[9050-76-4]<br>From <i>E. coli</i><br>RNase H is an endonuclease that cleaves the RNA of<br>RNA-DNA hybrids, producing a 3'-hydroxyl and a 5'-<br>phosphate at the cleavage point. However, RNase H does<br>not hydrolyze either RNA-RNA or DNA-DNA hybrids, or<br>single-stranded RNA or DNA.<br><b>Unit Definition:</b> One unit is the amount of enzyme<br>required to produce 1 nmole of acid-soluble<br>ribonucleotides from [ <sup>3</sup> H]poly(A) ù poly(dT) in 20 min, at<br>37°C<br>Our RNase H has been tested for contaminating DNase,<br>endonuclease, RNase III and non-specific RNase.<br>Offered in a 25 mM HEPES buffer, pH 8.0, 50 mM KCl, 1<br>mM DTT in 50% glycerol.<br><b>Ref.:</b> Donis-Keller, H., Nucleic Acids Res., <b>1</b> , 179 (1979);<br>(2) Okayama, H., Berg., P., Mol. Cell. Biol., <b>2</b> , 161 (1982). | 25 U<br>100 U  | <ul> <li>SP6/T7 Polymerase</li> <li>SP6/T7 Primers</li> <li>\alpha.<sup>32</sup>P Ribonucleotides</li> <li>Ultra-Pure Reagents</li> <li>Linbro@ MicroTubes</li> <li>Cloning Vectors</li> <li>Restriction Enzymes</li> <li>Buffers</li> <li>And Much More</li> </ul>  |               |
|                          |  |  | <ul> <li>T<sub>7</sub> RNA POLYMERASE</li> <li>T<sub>7</sub> RNA Polymerase is a DNA dependent RNA polymerase which has specificity for T<sub>7</sub> phage promoters. This allows the enzyme to efficiently synthesize <i>in vitro</i> transcripts from almost any DNA that is downstream from a T<sub>7</sub> promoter.</li> <li>Unit Definition: One unit is the amount of enzyme required to incorporate 1 nmol of labelled UTP into acid-soluble material in 60 min at 37°C.</li> <li>Supplied in a 50 mM Tris-HCI buffer, pH 7.9, 0.1M NaCI, 0.1 mM CDTA 1.0 mM diffusiterial and CO2 changed.</li> </ul>  | 5 KU<br>25 KU |
| 101079 <b>I</b><br>0-5°C | <b>RIBONUCLEASE T</b> <sub>1</sub><br>[9026-12-4]<br>From <i>Aspergillus oryzae</i> ,<br>E.C.3.1.27.1<br>Suspension in 0.70 saturated ammonium sulfate. Highly<br>purified.<br><b>Activity:</b> >300,000 units/mg.<br><b>Unit Definition:</b> One unit will produce acid soluble<br>oligonucleotides to cause a $\triangle A_{260}$ of 1.0 at pH 7.5 and<br>$37^{\circ}$ C.  | 100 KU<br>500 KU   | 0.1 min ED1A, 1.0 min dimittifetito and 50% glycerol.<br><b>Ref.:</b> (1) Chamberlin, M., and Ring, J., J. Biol. Chem., <b>248</b> , 2235-2244 (1973); (2) Tabor, S., and Richardson, C.C., Proc. Natl. Acad. Sci. USA, <b>82</b> , 1074-1078 (1985).  |               |
|                          |  |  | CLONING AND EXPRESSIONS VECTO  | ORS           |
|                          |  |  | AURORA™ AP<br>-20°C BASIC REPORTER VECTOR<br>Reporter vector (pSEAP) for expressing Secreted Alkaline<br>Phosphatase (SEAP) in mammalian cells.<br>Lacking eukaryotic promoter and enhancer sequences, it<br>serves as either a negative control or cloning vehicle for  | 20 µg         |
| ę                        | SP6 RNA POLYMERASE<br>Isolated from SP6 phage-infected Salmonella typhimurium.<br>very specific for the SP6 phage promoter sequences in DN<br>promoter sequences, when inserted into cloning vectors, ca<br>synthesize refined RNA transcripts from the cloned DNA se<br>transcripts are useful as hybridization probes and substrate  | The enzyme is<br>A. These<br>an be used to<br>equence. These<br>as for RNA | strong promoters. The SEAP gene is followed by the SV40 late polyadenylation and transcription pause sites (reduces background transcription). An f1 origin of replication allows for single-stranded DNA production. The pUC19 origin and ampicillin resistance gene allow for propagation and selection in <i>E. coli</i> .  |               |
|                          | processing.<br><b>Unit Definition:</b> One unit incorporates 1.0 nmol ATP into an acid soluble form in one hour at 37°C.   |  | 3131040 AURORA™ AP<br>-20°C CONTROL REPORTER VECTOR  | 20 µg         |

#### -20°C

#### AP Control Vector with the SV40 early promoter inserted upstream and enhancer inserted downstream of the SEAP gene. The promoter sequence contains the SV40 origin of

replication useful in cells expressing the T-antigen of SV40. It is useful as a positive control or as a reference for comparing the activities of promoters and enhancers.

Shipped on dry ice.

800709

800710

Shipping & Storage: Solution in 100 mM NaCl, 50 mM Tris-HCl, pH 7.9, 1 mM EDTA, 20 mM 2-mercaptoethanol, and 50% glycerol.

**Molecular Biology** 

200 U

1 KU

# **DNA Plasmid Vectors**



10 µg

10 µg

CATALOG NUMBER

### **DNA PLASMID VECTORS**

| AURORA™ AP<br>ENHANCER REPORTER VECTOR<br>AP Enhancer Vector with the SV40 enhancer downstream   | 20 µg   | DNA PLASMID VECTORS   |  |
|--|---|---|--|
| of the SEAP gene.<br>Ideal for the study of promoters cloned into the multiple<br>cloning sites. Potentially, the SV40 enhancer can increase<br>the transcriptional activity of weak promoters inserted in<br>the MCS.   |   | 821220 LAMBDA gt10 VECTOR<br>λgt 10 is a 43.34 Kb insertion vector with a unique EcoF<br>I restriction site within the repressor (cl) gene. It can<br>accept restriction fragments up to approximately 7.5 Kb in<br>lenoth, such as cDNA molecules with EcoR I ends, λqt 10   |  |
| AURORA™ AP<br>PROMOTER REPORTER VECTOR<br>AP Basic Vector that includes the SV40 early promoter (no<br>enhancer) inserted upstream of the SEAP gene.<br>The promoter fragment includes an SV40 origin of<br>replication for expression in cells that actively express the<br>SV40 T-antigen. pSEAP Promoter Vector may be used to<br>analyze enhancer sequences cloned into one of the<br>unique sites in the vector.  | 20 µg   | does not require an insert to be packageable and is thus<br>much more efficient at cloning smaller fragments, such as<br>cDNA molecules of 400-1500 base pairs, than other<br>related lambda insertion vectors.<br>$\lambda gt$ 10 is supplied as an aqueous solution in 10 mM tris<br>HCl, pH 8.0, 1 mM EDTA at a concentration of 0.5 $\mu g/\mu l$ .   |  |
| 0921100 <mark>AcMNPV C6</mark><br>-20℃ Wild Type Virus   | NPV C6     2 ml     821221     LAMBDA gt11 VECTOR       d Type Virus     λgt 11 is a 43.70 Kb expression vector caunique EcoR I restriction site within lacZ ge |   |  |
| <ul> <li>Destruction</li> <li>Destruct</li></ul> | 15 μg   | 53 bp upstream from the termination codon. Open readin<br>frames within the inserted DNA fragments may be<br>expressed as fusion proteins with the lacZ gene product,<br>galactosidase. Libraries constructed in $\lambda$ gt 11 can be<br>screened using antibody probes that recognize antigens<br>specified by the insert DNA. The functional $\beta$ -<br>galactosidase of gt 11 is usually inactivated by insertion of<br>foreign DNA and the percentage of recombinants can<br>pacified by the acar but the relative number of but and algor  |  |
| 0910100 BAC-UP6 VIRAL DNA<br>-20°C Derivative of AcMNPV that facilitates the production of<br>recombinant viral expression vectors from AcMNPV<br>transfer vectors.<br>A transfection reagent is provided with Bac-Up6 DNA for<br>efficient co-transfections.  | 15 μg   | plaques when libraries are plated in the presence of X-gal<br>and IPTG.<br>λgt 11 is supplied as an aqueous solution in 10 mM tris<br>HCl, pH 8.0, 1 mM EDTA at a concentration of 0.5 μg/μl.<br>ICN Reagents for Lambda Library Screening:   |  |
| <b>DBAC-UP8</b><br>Transfer vector for high-level expression of cloned genes<br>under the control of the potent AcMNPV polyhedrin<br>promoter.<br>Complete construct information is available upon request.  | 15 μg   | <ul> <li>Protein-A, [<sup>125</sup>]</li> <li>Biotrans™ PVDF</li> <li>Petri Plates</li> <li>Ampicillin</li> <li>Agar</li> <li>A tribodias</li> </ul>  |  |
| <ul> <li>D910300 BAC-UP9</li> <li>Transfer vector for high-level expression of cloned genes under the control of the potent AcMNPV polyhedrin promoter.</li> <li>Complete construct information is available upon request.</li> </ul>  | 15 μg   | • IPTG  |  |
| <ul> <li>BAC-UP BACULOVIRUS<br/>EXPRESSION KIT<br/>The complete kit includes the following material, sufficient<br/>to perform 5 transfections:</li> <li>2 Different transfer vectors for high-level expression of<br/>cloned genes driven by the potent AcMNPV polyhedrin<br/>promoter.</li> <li>Bsu36 I viral DNA digest.</li> <li>Transfection reagent.</li> <li>IPLB-Sf21 Spodoptera frugiperda cells.</li> <li>Positive control virus stock.</li> <li>Negative control AcMNPV wild-type virus.</li> <li>Bac1 sequencing/PCR Primer.</li> <li>Bac2 sequencing/PCR Primer.</li> <li>Positive control plasmid pBac-up8-GUS.</li> </ul>   | 1 kit   | <ul> <li>LAMBDA PHAGE DNA</li> <li>Produced from a <i>C1857 S7 Lysogen of E. coli</i> 200-400 μg. as determined by absorbancy at 260 nm.</li> <li>One OD<sub>260</sub> unit is approximately 50 micrograms of Lambda DNA. Optical density ratios are used as a criterior of purity, with a typical analysis giving A<sub>250</sub>/A<sub>260</sub> = .89 at pH 7.0 and A<sub>250</sub>/A<sub>260</sub> = .52 at pH 7.0.</li> <li>The viral DNA is extracted by a modification of the phenol method described by Kaiser and Hogness.</li> <li>Ref.: 1. Hedgpeth, J., Goodman, H.M., and Boyer, H.W., Proc. Nat. Acad. Sci. USA, 69, 3448 (1972).</li> <li>2. Kaiser, A.D., and Hogness, D.S., J. Molec. Biol., 2, 392 (1960).</li> </ul> |  |
| 0921000<br>-20°C   | 2 ml  |   |  |

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500µg 2mg