CATALOG NUMBER

153451 0-5°C **Xho I LINKER** 1 U
d(CCTCGAGG)
1u = 1 A_{260} unit. One A_{260} unit is approx. 40 µg of linker.

153452 0-5°C **Xho I LINKER** 1 U
d(CCCTCGAGGG)
1u = 1 A_{260} unit. One A_{260} unit is approx. 40 µg of linker.

153453 0-5°C **Xho I LINKER** 1 U
d(CCGCTCGAGCGG)
1u = 1 A_{260} unit. One A_{260} unit is approx. 40 µg of linker.

153454 0-5°C **Xma III LINKER** 1 U
d(CCGGCCGG)
1u = 1 A_{260} unit. One A_{260} unit is approx. 40 µg of linker.

DNA Primers

Primers are synthetic, single-stranded oligodeoxyribonucleotides used to prime (promote) DNA synthesis via DNA Polymerase and/or Reverse Transcriptase. Primers are also used in the generation of a hybridization probe for cloned sequences. Each product is tested for purity and performance.

153455 0-5°C **pBR322 *Bam*H I SITE, 20MER** 3 µg
(Clockwise)
5F128é...CACTATCGACTAC-GCGATCA...3′

153456 0-5°C **pBR322 *Eco*R I SITE, 16MER** 3 µg
(Clockwise)
5′...GTATCACGAGGCCCTT...3′

153457 0-5°C **pBR322 *Bam*H I SITE, 16MER** 3 µg
(Counter-clockwise)
5′...ATGCGTCCGGCGTAGA...3′

153458 0-5°C **pBR322 *Eco*R I SITE, 15MER** 3 µg
(Counter-clockwise)
5′...GATAAGCTGTCAAAC...3′

153459 0-5°C **pBR322 *Hind* III SITE, 15MER** 3 µg
(Clockwise)
5′...GACAGCTTATCATCG...3′

153460 0-5°C **pBR322 *Hind* III SITE, 16MER** 3 µg
(Counter-clockwise)
5′...GCAATTTAACTGTGAT...3′

153461 0-5°C **pBR322 *Pst* I SITE, 16MER** 3 µg
(Clockwise)
5′...GCTAGAGTAAGTAGTT...3′

153462 0-5°C **pBR322 *Pst* I SITE, 15MER** 3 µg
(Counter-clockwise)
5′...AACGACGAGCGTGAC...3′

153463 0-5°C **pBR322 *Sal* I, 15MER** 3 µg
(Clockwise)
5′...ATGCAGGAGTCGCAT...3′

153464 0-5°C **pBR322 *Sal* I, 15MER** 3 µg
(Counter-clockwise)
5′...AGTCATGCCCCGCGC...3′

CATALOG NUMBER

821335 **gt 11 FORWARD** 2 µg
5′-GACTCCTGGAGCCCG-3′
These primers are complementary to parts of the lacZ gene of gt 11 at either side of the Eco RI cloning site. Their use enables direct dideoxy sequencing of each end of the inserted DNA without further subcloning.

821336 **gt 11 REVERSE** 2 µg
gt 11 reverse
5′-GGTAGCGACCGGCGC-3′
These primers are complementary to parts of the lacZ gene of gt 11 at either side of the Eco RI cloning site. Their use enables direct dideoxy sequencing of each end of the inserted DNA without further subcloning.

HIV Primers
See: HIV Products Section

821228 **M13 15-MER** 2 µg
5′-TCCCAGTCACGACGT-3′.
This was the first M13 sequencing primer developed and it anneals to the (+) strand 37 bases upstream of the first restriction site in the multiple cloning site.

821229 **M13 SEQUENCING PRIMER (-20) 17-MER** 4 µg
5′-GTAAAACGACGGCCAGT-3′.
A universal primer with little complimentarity to other regions of the M13 DNA. It anneals close to the multiple cloning site. Since pUC vectors contain the same β-galactosidase gene as M13mp18/19, the same primer can be used to determine DNA sequences cloned into pUC vectors.

153465 0-5°C **M13 SEQUENCING PRIMER (-40) 17-MER** 4 µg
5′...GTTTTCCCAGTCACGAC...3′

153466 0-5°C **M13 HYBRIDIZATION PROBE PRIMER, 16-MER** 3 µg
5′...CACAATTCCACACAAC...3′

821230 **M13 PROBE PRIMER** 2 µg
5′-GAAATTGTTATCC-3′.
This primer is used
to generate hybridization probes from (+) strand M13 DNA carrying inserts.
The primer hybridizes downstream of the cloning site and during second strand synthesis in the presence of radioactive precursors, a partially double stranded molecule is generated which is single stranded in the region of the inserted DNA. This region of the inserted DNA can base pair with complementary sequences during hybridization reactions.

821231 **M13 REVERSE SEQUENCE PRIMER** 2 µg
5′-AACAGCTATGACCATG-3′
This reverse sequencing primer can be used to obtain sequence from the opposite end of the
insert in M13/pUC vectors to that obtained using the universal M13 primer.
It is not complementary to the (+) strand of M13 phage and therefore either double stranded sequencing must be performed or a partial second strand is generated using Klenow enzyme.

CATALOG NUMBER

821237 OLIGO (dC)-ECOR I — 2 µg
5′-TGAATTCGGATCCCC CCCCCCCCC-3′.
This oligonucleotide is used to prime the synthesis of a second strand when the first strand has been tailed with oligo d(G). After repair and if the first strand has been primed using the oligo d(T)-Eco RI.

821338 OLIGO (dG)-ECOR I — 2 µg
5′-GTGAATTCGTCGAC GGGGGGGGGG-3′.
This primer is an alternative to the oligo d(C)-Eco RI primer allowing tailing of the first strand with oligo d(C). Additionally, the Bam HI site is replaced by a Sal 1 site.

821339 OLIGO (dT)-ECOR I — 2 µg
5′-TGAATTCTTTTTTTTTTTTTTTT-3′.
This primer is used in place of oligo d(T) in the first strand synthesis reaction when making cDNA. On completion of the second strand an Eco RI site is present at the end of the molecule.
All oligonucleotides are supplied in aqueous solutions in 10 mM tris HCl, pH 8.0, 1 mM EDTA at a concentration of 1 µg/ml.

821232 SP6 PRIMER — 2 µg
5′-ACCTTATGTATCATACACAT-3′.
The SP6 primer hybridizes to part of the SP6 promoter found in many recently developed transcription vectors. It allows dideoxy sequencing of the transcribed insert DNA in such vectors.

153467 0-5°C SP6 PROMOTER PRIMER, 17-MER — 3 µg
5'...ATTTAGGTGACACTATA...3'

821233 T3 PRIMER — 2 µg
T3 primer is complementary to part of the T3 promoter and is used to sequence part of the insert DNA in transcription vectors containing the T3 promoter.

821234 T7 PRIMER — 2 µg
5′-CTCACTATAGGGAGACC-3′. This primer is complementary to conserved sequences present in T7 promoters and permits dideoxy sequencing of DNA in transcription vectors containing T7 promoters.

153468 0-5°C T7 PROMOTER PRIMER, 17-MER — 3 µg
5'...TAATACGACTCACTATA...3'

MOLECULAR BIOLOGY CELL CULTURE COMPONENTS

194021 RT AGAR, Bacteriological — 250 g, 1 kg
[9002-18-0]
Molecular Biology Reagent
Specially purified for use in preparing solid culture media for microbiological and bacteriological applications. Naturally occurring impurities have been reduced to a minimum.

194022 RT AGAR — 250 g, 1 kg
[9002-18-0]
Molecular Biology Reagent
Powder
Suitable as a component in culture media for molecular genetics.

CATALOG NUMBER

194023 RT D-(-)-ARABINOSE — 25 g, 100 g
[28697-53-2]
Molecular Biology Reagent
Purity: 99%
Suitable as a culture media component.
$C_5H_{10}O_5$ MW 150.1

194028 RT 2-DEOXY-D-GLUCOSE — 250 mg, 1 g, 5 g
[154-17-6]
White crystals
Purity: 99%
For use as a culture media component for molecular genetics.
$C_6H_{12}O_5$ MW 164.2

194024 RT D-(+)-GLUCOSE — 250 g, 1 kg
[50-99-7]
(Dextrose)
Molecular Biology Reagent
Ideal as a culture media component.
$C_6H_{12}O_6$ MW 180.2

193996 RT GLYCEROL — 100 ml, 500 ml, 1 liter
[56-81-5]
Molecular Biology Reagent
Purity: 99+%
Heavy metals (Pb): <5 ppm
No detectable DNase, RNase, or protease.
Prevents back-diffusion and protein samples into the buffer.
$C_3H_8O_3$ MW 92.09

194025 0-5°C HEMIN — 1 g, 5 g
[15489-47-1]
(Hemin Chloride)
Molecular Biology ReagentSource: Bovine
Ideal for use in culture media for molecular genetics.
$C_{34}H_{32}ClFeN_4O_4$ MW 652

102040 RT INDOLE-3-ACRYLIC ACID — 1 g, 5 g, 10 g
[1204-06-4]
Crystalline
Light yellow crystals.
A metabolite of tryptophan.
$C_{11}H_9NO_2$ MW 187.2

194029 0°C ISOPROPYL-β-D-THIOGALACTOPYRANOSIDE — 100 mg, 250 mg, 500 mg, 1 g, 5 g, 10 g
[367-93-1]
(Isopropyl-β-D-Thiogalactoside; IPTG)
Molecular Biology Reagent
Purity: >99%
Cell culture media component for use in molecular genetics.
β-Galactoside inducer.
$C_9H_{18}O_5S$ MW 238.3

194026 0°C β-NICOTINAMIDE ADENINE DINUCLEOTIDE — 500 mg, 1 g
[53-84-9]
(β-NAD)
Molecular Biology Reagent
Purity: ~99%
For use as a culture media component for molecular genetics. Chromatographically purified to remove trace inhibitors.
$C_{21}H_{27}N_7O_{14}P_2$ MW 663.4